

*Research Article***Immunoexpression of Stat γ in Human Fetal Ovary During the Second Trimester Pregnancy.****G. M. Hartshorne***, **Mohamed M. Ahmed****, **Mostafa Abdel-Hamid**** and **Sabreen M. Ahmed****.

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Abstract

Background: In the development of the human fetal ovary, the second trimester includes the transition from oogonial replication to primordial follicle formation. The STAT family proteins plays an important role during germ cells differentiation in the human fetal ovary. The study was conducted at Warwick medical school research institute, UK. to assess differentiation of germ cells in a series of female gonads from the second trimester using STAT γ immunofluorescence technique. **Materials and Methods:** Human fetal ovaries were obtained from second trimester fetuses (14 to 24 weeks gestation) after induced abortion. Then the tissues were cut into Pieces for processing for paraffin sections. Ovarian pieces were studied for the presence of germ cell marker using immunohistochemistry for STAT γ . **Results:** STAT γ was expressed in the cytoplasm of oocytes during oogenesis and folliculogenesis in human fetal ovary. While there was no expression in somatic cells. When STAT γ was activated, it moved from the cytoplasm to the nucleus. **Conclusions:** STAT γ is a useful tool for germ cell studies; the expression of STAT γ in human fetal ovary enable one to follow oogenesis throughout the second trimester pregnancy. So, the present study will serve as a reference for further analyses investigating the developing human fetal ovary.

Key words: STAT γ , Human fetal ovary, Second trimester.**Introduction**

STAT γ is one of a series of cytosolic STAT (signal transducer and activator of transcription) proteins known to be involved in transcriptional regulation.⁽¹⁾ STAT proteins are activated by hormones, growth factors and extracellular signaling proteins such as cyto-kines.⁽¹⁾ The STAT members are seven proteins (STAT 1, 2, 3, 4, 5A, 5B, and 6) which are identified in mammals and all are important for cellular signaling.⁽¹⁾ Activated STAT γ is required for dimerization, nuclear translocation and transcriptional activity.⁽²⁾

STAT γ has a critical role in different biological functions such as cell growth, survival and motility depending on the stimulus and the cell type.⁽²⁾ It is essential during embryogenesis and its disruption is embryonic lethal.⁽³⁾ Also, during implantation and decidualization, it is important for mouse and rat reproduction.⁽⁴⁾ Additionally, leptin regulates ovarian granulosa cell function through JAK/STAT pathway. STAT γ pathway

is involved in mediating epidermal growth factor (EGF) which stimulates ovarian follicular development.⁽⁵⁾

STAT γ protein is expressed in the cytoplasm of oocytes from primordial, primary and secondary follicles in the adult ovary and in developing acrosomes of round spermatids in the adult testis. STAT γ is also expressed in gonocytes, prospermatogonia and oocytes of embryonic and neonatal gonads.⁽³⁾

The present study was carried out to assess differentiation of germ cells in a series of female gonads from the second trimester. The oocyte marker STAT γ , has been used because it played an essential role in gem cell differentiation in mice.

Materials and Methods**Tissue samples:**

The present study was approved by Warwickshire Research Ethics Committee UK.

Human fetal ovaries were obtained from second trimester fetuses (17 to 24 weeks gestation) after induced abortion. None of the terminations were for reasons of fetal abnormality. Gestational age was estimated through ultrasound examination before termination. Ovaries were dissected carefully from the fetal abdominal cavity and placed in collection medium for preservation.⁽¹⁾

The human fetal ovaries were cut into uniform sized wedges (3x 3x3 mm). Freezing and thawing of ovarian tissue was done according.⁽¹⁾ The ovarian fragments were transferred into cryovials (one fragment per vial) containing 1ml of a cryopreservation medium. The slow freezing procedure was performed in a programmable controlled rate freezing machine for two hours then all the vials

were plunged into liquid nitrogen tank at -196°C.

Pieces of human fetal ovaries were processed for paraffin sections.⁽¹⁾ To assure satisfactory quality, tissue samples were assessed by hematoxy-lineos in staining. Any poor sample was excluded from further analysis.

Fluorescent immunocytochemical staining

Human fetal ovarian pieces were studied for the presence of germ cell marker using immunofluorescence for STAT3. The primary and secondary antibodies used are indicated in Table (I). Negative control was performed by omitting the primary antibody which was replaced by normal serum resulting in complete absence of signals and no staining was observed in this control.

Table (1): The primary and secondary antibodies for STAT3.

Primary antibody	Company	Pre-treatment	Dilution	Secondary antibody	Dilution	Company
STAT3 (C30) Rabbit anti-human	Santa cruz, UK	Enzymatic	1:500	Goat anti-rabbit IgG FITC	1:1000	Sigma, UK

Statistical analysis

Data were presented as mean±standard deviation (SD). Statistical significance was determined by unpaired student’s t-Test of unequal variance. The relationship between age (gestation weeks) and the number of positive STAT3 stained cells was investigated using pearson correlation coefficient. Results were considered significant at p<0.05.

Results

From 17 weeks to 24 weeks the occurrence of STAT3 immunofluorescence in human oocytes was detected by the use of high power view of a scanning laser confocal microscopy. Negative controls were shown in (Figure 1).

STAT3 was not detected at 18 weeks gestation but began to be expressed at 19 weeks gestation in developing human ovary.

STAT3 protein immune-fluorescence was seen in the cytoplasm of all oocytes within oocytes nests (Figure 2), in early follicles and in primary follicles at the interface with the follicle cells

(Figure 3). Oocytes in early follicles after nest breakdown were clearly stained for STAT3 in the cytoplasm, but there was no labelling of somatic stromal or granulosa cells with STAT3 (Figure 3, 4 and 5). Cytoplasm was stained for STAT3 at similar intensity for small and growing oocytes (Figure 2, 3 and 5). STAT3 became progressively stronger in the cytoplasm of oocytes as age advanced while there was no expression in somatic cells.

The stained sections confirmed that STAT3 is a germ cell (oocyte) marker in humans (Figure 5). When STAT3 is activated, it moves from the cytoplasm to the nucleus. At 24 weeks gestation, there was a strong nuclear staining (Figure 4).

Figure 6 summarizes the results of a quantitative assessment of the STAT3 marker at different gestational ages (weeks).

There was a strong positive correlation between the age (gestation weeks) and the number of STAT3 positive stained cells (Table 2).

There was a statistically significant difference at the P < 0.05 level in the number of STAT3

positive stained cells in relation to age between the eight groups ($P=0.003$) (Table 3).

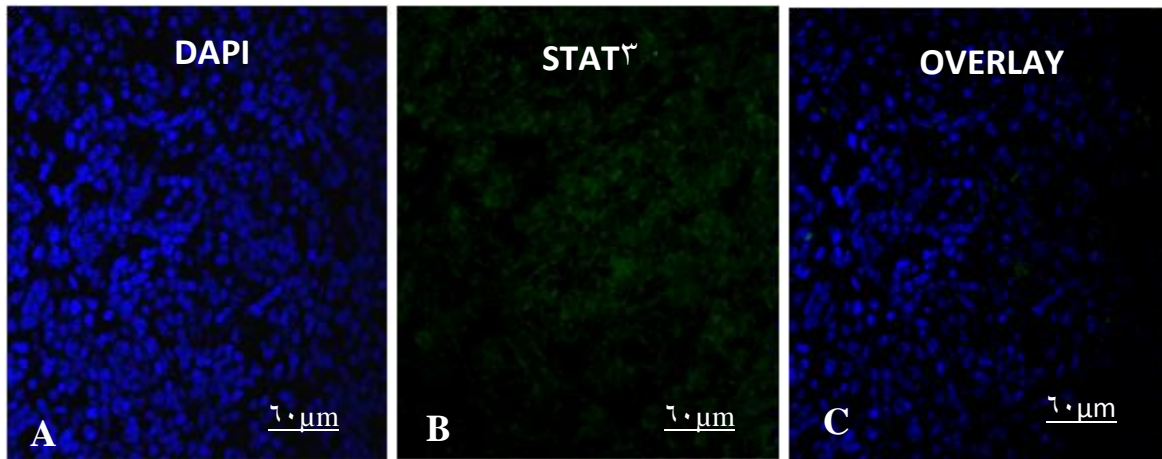


Figure (1): Photomicrographs of a section in fetal ovary (gestational age 19 weeks) showing negative control for STAT3. (A) Shows confocal immunostained section with DAPI as a nuclear stain, (B) Shows no signal in absence of primary antibody for STAT3 and (C) Shows overlay.

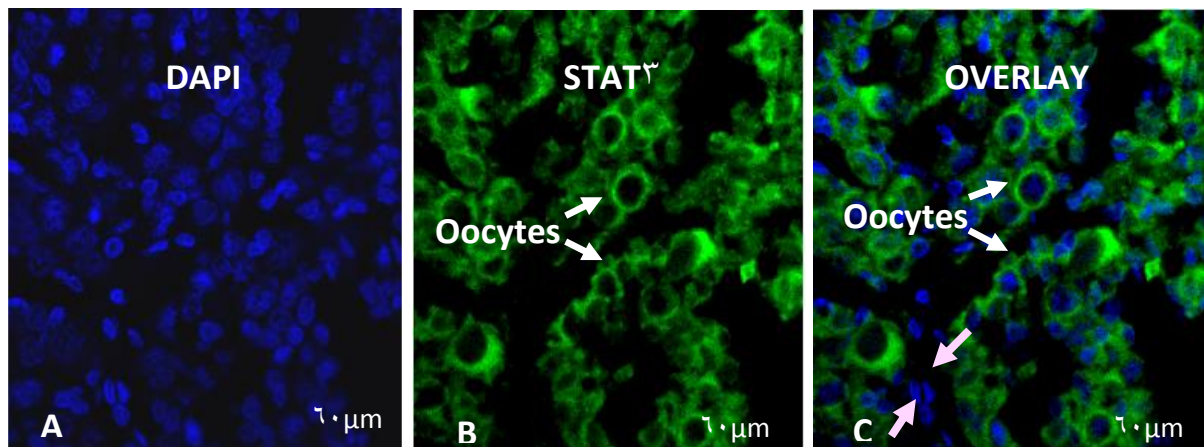


Figure (2): Photomicrographs of a section in fetal ovary (gestational age 21 weeks). Arrows in (B) and (C) indicate oocytes with cytoplasmic labeling. Arrows in (C) indicate unstained somatic cells.

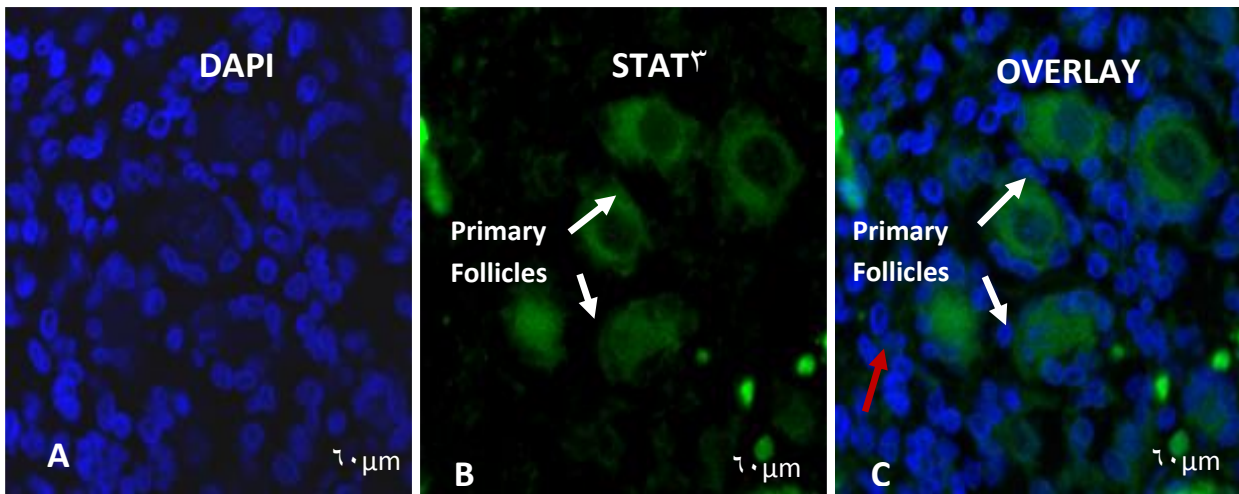


Figure 3: Photomicrographs of a section in fetal ovary (gestational age 27 weeks). Arrows in (B) and (C) indicate cytoplasmic labeling of oocytes enclosed in follicle cells, red arrow indicates absence of labeling in granulosa cells

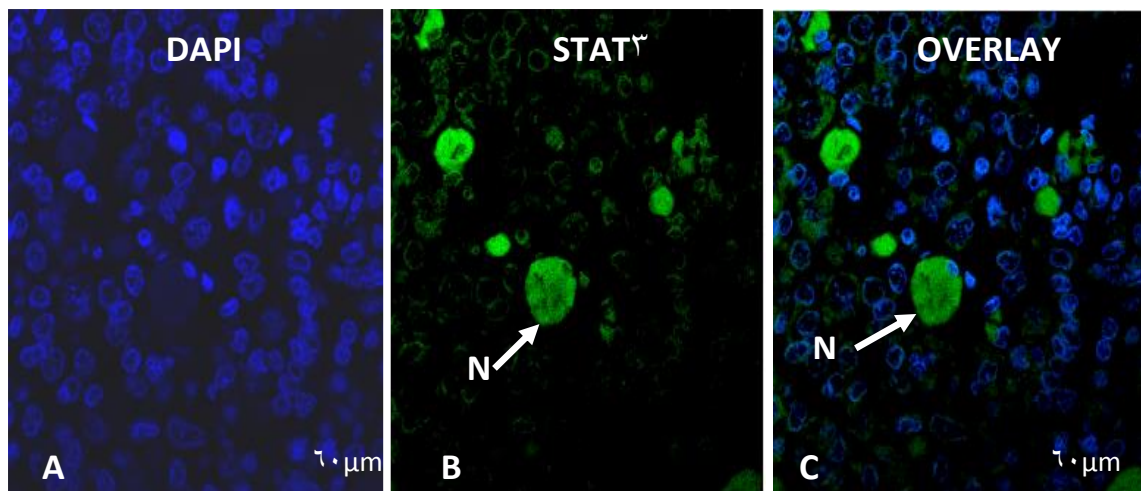


Figure 4: Photomicrographs of a section in fetal ovary (gestational age 24 weeks). Arrows in (B) and (C) indicate strong nuclear labeling after STAT3 protein activation and movement from oocyte cytoplasm to oocyte nucleus.

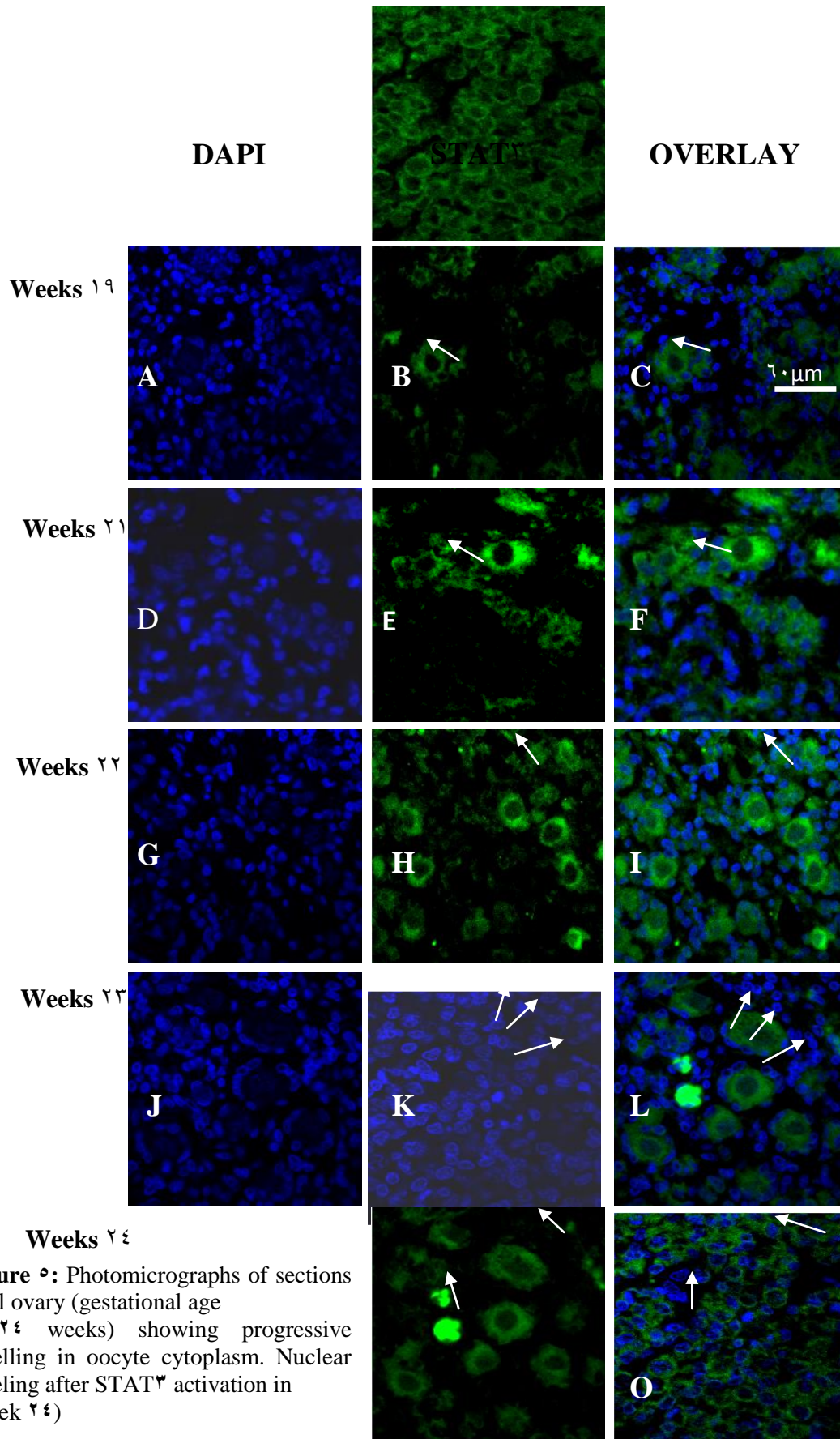


Figure 9: Photomicrographs of sections fetal ovary (gestational age 19-24 weeks) showing progressive labelling in oocyte cytoplasm. Nuclear labeling after STAT γ activation in (week 24)

in

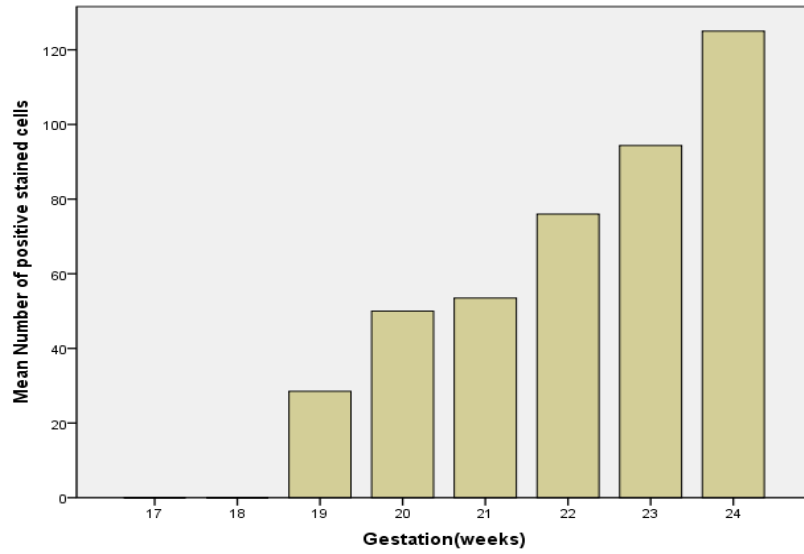


Figure 6: The mean number of STAT3 positive stained cells at different gestational ages (weeks).

Table 7: The relationship between age (gestation weeks) and the number of STAT3 positive stained cells using pearson correlation coefficient.

		Gestation(weeks)	Number of positive stained cells
Gestation(weeks)	Pearson Correlation	1	.893**
	Sig. (2-tailed)		.000
	N	49	49
Number of positive stained cells	Pearson Correlation	.893**	1
	Sig. (2-tailed)	.000	
	N	49	49

Table 8: A one way analysis of variance (ANOVA) between age groups to explore the impact of age on the number of STAT3 positive stained cells. Significance level ($p \leq .05$ level).

	Sum of Squares	Df	Mean Square	F	Sig.
Between Groups	22.71.707	7	3103.108	7.969	.003
Within Groups	4976.870	11	452.443		
Total	27.048.632	18			

Discussion

In the present study, human oocytes showed STAT γ protein immunofluorescence in the cytoplasm of all oocytes within oocytes nests, early follicles and primary follicles at the interface with the follicle cells, oocytes in early follicles after nest breakdown were clearly stained for STAT γ in the cytoplasm. Cytoplasm was stained for STAT γ at similar intensity for small and growing oocytes. This is in agreement with Wen et al. (2006)⁽⁴⁾ who demonstrated that STAT γ is expressed in high levels in ovary.

In agreement with Antczak and Blerkom⁽¹⁷⁾ and Murphy et al.⁽¹⁸⁾, there was no labelling of somatic stromal or granulosa cells with STAT γ . Explanation of this notion is that STAT γ interact with oocyte cytoplasmic receptor like molecule to involve a signal transduction pathway that is somewhat different from the Jak-STAT pathway in somatic cells.⁽¹⁾

STAT γ was not detected at 18 weeks gestation but began to be expressed at 19 weeks gestation in developing human ovary. Murphy et al.⁽¹⁸⁾; Hou et al.⁽¹⁴⁾ reported that the immunoexpression of STAT γ was not detected at 12.5 day post coital (dpc) but began at 13.5 (dpc) in developing ovaries of mice. These periods are equivalent to the second trimester pregnancy in human.

In the present study, in the developing ovary, STAT γ became progressively stronger in the cytoplasm of oocytes while there was no expression in somatic cells. This is supported by the findings of Murphy et al. and Hou et al. in mice^(4, 14).

When STAT γ is activated, it moves from the cytoplasm to the nucleus. At 24 weeks gestation, there was a strong nuclear staining. Therefore, STAT γ was activated in some germ cells. Up to the researcher knowledge, this finding was not supported by previous research. Murphy et al.⁽⁴⁾ did not observe nuclear staining at any examined mice ovary. Therefore, STAT γ was not active in mice germ cells.

Antczak and Blerkom⁽¹⁷⁾ explained the process of STAT γ activation; Cytoplasmic STAT γ proteins are phosphorylated by a Jak tyrosine kinase following the activation of a particular set of receptors. Phosphorylated STAT proteins

form homo- or heterodimers and rapidly translocate to the nucleus to initiate specific gene transcription. Also, Matsuoka et al.⁽¹⁹⁾ results suggest possible roles of leptin in several aspects during oocyte maturation by activating the STAT signal transduction pathway.

In the present study, there was a strong positive correlation between the gestational age and the numbers of STAT γ positive cells. This finding is in agreement with Murphy et al.⁽⁴⁾

It could be concluded that STAT γ is a useful tool for germ cell studies; the expression of STAT γ in human fetal ovary enable one to follow oogenesis throughout the second trimester pregnancy. So, the present study will serve as a reference for further analyses investigating the develop-ping human fetal ovary.

Since, human fertility depends upon a supply of oocytes (eggs) being available for ovulation during the adult life of the woman. The present study will add to other studies that aim to find out the factors that control the ovarian reserve of oocytes that last throughout the female reproductive period. This might help in solving female infertility by finding novel ways that ensure sound oocyte supply to be fertilized.

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